

Inhibition of adipose tissue lipolysis increases intramuscular lipid use in type 2 diabetic patients

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Inhibition of adipose tissue lipolysis increases intramuscular lipid use in type 2 diabetic patients

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Abstract *Aims/hypothesis:* In the present study, we investigated the consequences of adipose tissue lipolytic inhibition on skeletal muscle substrate use in type 2 diabetic patients. *Materials and methods:* We studied ten type 2 diabetic patients under the following conditions: (1) at rest; (2) during 60 min of cycling exercise at 50% of maximal workload capacity and subsequent recovery. Studies were done under normal, fasting conditions (control trial: CON) and following administration of a nicotinic acid analogue (low plasma non-esterified fatty acid trial: LFA). Continuous [U-¹³C]palmitate and [6,6-²H₂]glucose infusions were applied to quantify plasma NEFA and glucose oxidation rates, and to estimate intramuscular triacylglycerol (IMTG) and glycogen use. Muscle biopsies were collected before and after exercise to determine net changes in lipid and glycogen content specific to muscle fibre type. *Results:* Following administration of the nicotinic acid analogue (Acipimox), the plasma NEFA rate of appearance was effectively reduced, resulting in lower NEFA concentrations in the LFA trial ($p<0.001$). Plasma NEFA oxidation rates were substantially reduced at rest, during exercise and subsequent recovery in the LFA trial.

The lower plasma NEFA oxidation rates were compensated by an increase in IMTG and endogenous carbohydrate use ($p<0.05$). Plasma glucose disposal rates did not differ between trials. In accordance with the tracer data, a greater net decline in type I muscle fibre lipid content was observed following exercise in the LFA trial ($p<0.05$). *Conclusions/interpretation:* This study shows that plasma NEFA availability regulates IMTG use, and that adipose tissue lipolytic inhibition, in combination with exercise, could be an effective means of augmenting intramuscular lipid and glycogen use in type 2 diabetic patients in an overnight fasted state.

Keywords Acipimox · IMCL · IMTG · Insulin sensitivity · Skeletal muscle

Abbreviations CON: control trial · FA: fatty acids · HOMA-IR: homeostasis model assessment insulin resistance index · IMTG: intramyocellular triacylglycerol · LFA: low plasma fatty acid trial · R_a : rate of appearance · R_d : rate of disappearance · R_{ox} : rate of oxidation · TG: triacylglycerol · $V_{O_{2\max}}$: maximal oxygen uptake capacity · W_{\max} : maximal workload

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Introduction

Numerous studies have reported the association between elevated plasma NEFA concentrations, intramyocellular triacylglycerol (IMTG, often also abbreviated to IMCL) accumulation and the development of insulin resistance and/or type 2 diabetes [1–7]. The Randle (glucose–fatty acid) cycle has often been used to explain the mechanism behind skeletal muscle insulin resistance induced by fatty acid (FA) [8]. More recent insights from various lipid infusion studies have led to an alternative mechanism [9–13]. The latter suggests that elevated NEFA delivery and/or impaired FA oxidation result in intramyocellular accumulation of triacylglycerol (TG) and FA metabolites (such as fatty acyl-CoA, ceramides and diacylglycerol), which

induce defects in the insulin signalling cascade, causing skeletal muscle insulin resistance [9, 10, 13–17]. Insulin resistance can subsequently lead to development of the hyperglycaemic and/or hyperinsulinaemic state that is associated with type 2 diabetes. The latter is accompanied by major disturbances in skeletal muscle substrate metabolism [18, 19], which further stimulate IMTG accretion at the expense of IMTG mobilisation and subsequent oxidation [20].

Therefore, excess IMTG accretion in sedentary, obese and/or type 2 diabetic patients is the result of a structural imbalance between NEFA uptake, IMTG deposition and its subsequent mobilisation and/or oxidation. Recent data indicate that this imbalance, and not the actual size or intramyocellular distribution of the IMTG pool, is responsible for the development of skeletal muscle insulin resistance [21–23]. It has been speculated that an elevated IMTG utilisation rate will prevent excess accumulation of intramyocellular TG and/or FA (metabolites). In addition, a greater IMTG turnover rate may reduce the degree of intramyocellular lipid peroxidation [17], due to a decreased residence time of the lipid deposits. The turnover of the IMTG pool should, therefore, form a major therapeutic target in the prevention and/or treatment of skeletal muscle insulin resistance, and interventions should be developed that stimulate IMTG use [24].

Exercise has been shown to augment IMTG use in healthy, fit males and probably represents an effective way to prevent and/or reduce excess IMTG accretion [24, 25]. Recently, we observed that even a single bout of endurance exercise can reduce type I muscle fibre lipid content by more than 60% in trained men [24, 25]. In the same study, we observed a progressive decline in IMTG oxidation rate over time during exercise, which was associated with a concomitant increase in plasma NEFA concentration [25]. The latter supported the contention that elevated plasma NEFA availability suppresses IMTG mobilisation [25–28]. Because obesity and/or type 2 diabetes are associated with elevated plasma NEFA concentrations [13, 29], it has been questioned whether IMTG stores are readily available as a substrate source in type 2 diabetic patients [24, 25]. Therefore, the first aim of the present study was to quantify IMTG use at rest, during exercise and subsequent recovery in type 2 diabetic patients. Various studies [24, 30–33] have reported that pharmacological inhibition of adipose tissue lipolysis can stimulate intramuscular lipid and/or glycogen use. Based on those findings, we questioned whether such an intervention could be used to lower plasma NEFA availability and increase the oxidation rate of the intramuscular lipid deposits at rest, during exercise and/or post-exercise recovery in type 2 diabetic patients.

This study provides novel insight into the role of plasma NEFA availability in modulating IMTG use in type 2 diabetic patients: It also suggests that adipose tissue lipolytic inhibition could be an effective means of augmenting intramuscular lipid and glycogen use.

Subjects, materials and methods

Subjects

Ten sedentary, overweight subjects, who had been diagnosed with type 2 diabetes for over 5 years, participated in this study (Table 1). All subjects were using oral blood-glucose-lowering medication (metformin with or without a sulphonylurea derivative). Type 2 diabetic status was verified with an OGTT according to WHO criteria [34]. In addition, insulin resistance was estimated using the homeostasis model assessment for insulin resistance index (HOMA-IR) [35]. Medication was withheld for 24 h prior to the experimental trials. Subjects were informed about the nature and risks of the experimental procedures before their written informed consent was obtained. This study was approved by the local Medical Ethical Committee.

Pre-testing

Maximal workload (W_{\max}) and oxygen uptake capacity ($V_{O_2 \max}$) were measured on a cycle ergometer (Lode Excalibur, Groningen, the Netherlands) during an incremental exercise test. Body composition was assessed using the hydrostatic weighing method. Body fat percentage was calculated using Siri's equation [36].

Diet and activity prior to testing

All subjects maintained normal physical activity patterns throughout the experimental period. In addition, they filled out a food intake diary for 2 days prior to the first trial

Table 1 Subjects' characteristics ($n=10$)

Age (years)	60.0±2.1
Height (m)	1.79±0.02
Body mass (kg)	90.6±3.0
BMI (kg/m ²)	28.4±1.0
Body fat percentage (%)	28.9±1.8
Fat free mass (kg)	64.0±1.9
Basal plasma glucose (mmol/l)	9.4±0.6
Plasma glucose _{120 min} (mmol/l)	16.7±1.2
Basal plasma insulin (mU/l)	9.2±1.0
HOMA-IR (mmol mU ⁻¹ l ⁻¹)	3.6±0.4
HbA _{1c} (%)	7.3±0.3
$V_{O_2 \max}$ (l/min)	2.9±0.2
W_{\max} (W)	200±15
Maximal heart rate (bpm)	161±4
Diagnosed with diabetes (years)	7.5±1

Body mass index (BMI) was calculated by dividing body mass by the square of the height (m). Insulin resistance was estimated by the homeostasis model assessment (HOMA-IR), which is calculated by dividing the product of basal plasma glucose and insulin concentrations by 22.5

Data represent means±SEM

in order to keep their dietary intake as identical as possible prior to the other trials. Dietary analyses showed a daily energy intake of 8.4 ± 0.7 MJ/day, with 55 ± 1 , 29 ± 1 and $16 \pm 1\%$ of energy intake derived from carbohydrate, fat and protein respectively. The evening before each trial, subjects received a standardised meal (41.5 kJ/kg of body weight; containing 58, 31 and 11% energy carbohydrate, fat and protein, respectively).

Experimental trials

Each subject performed three trials, separated by a 1-week interval. Each trial consisted of 90 min resting measurements, followed by 60 min cycling exercise ($50\% W_{\max}$) and 120 min of recovery. In the two main trials, a $[U-^{13}C]$ palmitate and $[6,6-^2H_2]$ glucose tracer were infused with breath, blood and muscle samples being collected at regular intervals. In the LFA trial, plasma NEFA availability was reduced by oral administration of a peripheral lipolytic inhibitor (Acipimox). In the CON trial a placebo was provided. A third trial was performed to determine the acetate recovery factor [37].

Protocol

After an overnight fast, subjects arrived at the lab at 08.00 hours, either by car or public transportation. After 30 min of supine rest, a percutaneous muscle biopsy was taken from the vastus lateralis muscle. A Teflon catheter was inserted into an antecubital vein of one arm for blood sampling; another catheter was inserted in the contralateral arm for isotope infusion. A resting blood and expired breath sample were collected (between 08.30 and 09.00 hours), after which an oral dose of 250 mg Acipimox or a placebo was administered ($t=0$ min). Exactly 30 min later ($t=30$ min), subjects were given a single intravenous dose of $NaH^{13}CO_3$ (0.06375 mg/kg), followed by a $[6,6-^2H_2]$ glucose prime (13.5 $\mu\text{mol/kg}$). Thereafter, a continuous infusion of $[6,6-^2H_2]$ glucose (0.3 $\mu\text{mol kg}^{-1} \text{min}^{-1}$) and $[U-^{13}C]$ palmitate (0.01 $\mu\text{mol kg}^{-1} \text{min}^{-1}$) (or $[1,2-^{13}C]$ acetate (0.08 $\mu\text{mol kg}^{-1} \text{min}^{-1}$) was started via a calibrated pump (IVAC, San Diego, CA, USA). At $t=150$ min, another dose of 250 mg Acipimox or a placebo was administered. At $t=120$ min, subjects started to exercise at $50\% W_{\max}$ for a 60-min period. While at rest, V_{O_2} and V_{CO_2} were measured continuously (Oxycon β ; Mijnhart, Bunnik, the Netherlands), during exercise V_{O_2} and V_{CO_2} were measured for 5 min every 15 min before sampling of blood and expired breath. Immediately after cessation of exercise, a second muscle biopsy was taken, after which the subjects rested in a supine position. V_{O_2} and V_{CO_2} were measured continuously during the second hour of post-exercise recovery ($t=240$ – 300 min).

Acipimox administration

Both at $t=0$ and 150 min, subjects were given 250 mg Acipimox (Nedios, Byk, Zwanenburg, the Netherlands) or a placebo. Acipimox is a potent adipose tissue lipolytic inhibitor and is generally prescribed for the treatment of hyperlipidaemia. Side-effects of Acipimox include: flushing, skin rashes, gastrointestinal complaints and headaches. In the present study, six out of ten subjects showed flushing (redness of the skin; predominantly in the face and on the back) about 30 min after ingesting the first dose. During exercise these effects were no longer distinguishable from the redness/flushing secondary to increased physical activity, as observed in the CON trial. Other side-effects were not reported. Subjects did not report a difference in perceived exertion between trials. In accordance, mean heart rate values during exercise, measured with a telemetric heart rate monitor (Polar Vantage XL; Polar, Kempele, Finland), did not differ between trials and averaged 116 ± 4 , 121 ± 3 and 122 ± 4 beats per min in the LFA, CON and acetate recovery trial, respectively.

Blood and breath sample analysis

Blood samples (7 ml) were collected in EDTA-containing tubes and centrifuged at $1,000 \times g$ for 10 min at 4°C . Aliquots of plasma were frozen immediately in liquid nitrogen and stored at -80°C . Plasma glucose (Roche, Basel, Switzerland), lactate, NEFA (Wako Chemicals, Neuss, Germany), glycerol (Roche Diagnostics, Indianapolis, IN, USA) and triglyceride (Sigma Diagnostics, St Louis, MO, USA) concentrations were analysed with a COBAS semiautomatic analyser (Roche). Plasma insulin was measured by radioimmunoassay (Linco, St Charles, MO, USA). Blood HbA $_{1c}$ content was analysed by high-performance liquid chromatography (Bio-Rad Diamat, Munich, Germany). Expired breath samples were analysed for $^{13}C/^{12}C$ ratio by gas chromatography continuous-flow isotope ratio mass spectrometry (MAT 252; Finnigan, Bremen, Germany). To determine plasma palmitate and NEFA kinetics, NEFA were extracted, isolated by thin-layer chromatography and derivatised to their methyl esters. Palmitate concentration was determined using analytical gas chromatography with flame ionisation detection using heptadecanoic acid as internal standard and comprised $21.3 \pm 0.7\%$ of total NEFA. Isotope tracer: trace ratios (TTR) of $[U-^{13}C]$ palmitate were determined using gas chromatography combustion isotope ratio mass spectrometry (Finnigan MAT 252). Following derivatisation, plasma $[6,6-^2H_2]$ glucose enrichment was determined by electron ionisation gas chromatography-mass spectrometry (Finnigan). Palmitate, glucose and acetate tracer concentrations in the infusates averaged 1.06 ± 0.01 , 34.5 ± 0.8

and 4.94 ± 0.02 mmol/l, respectively. Therefore, the exact tracer infusion rates averaged 9.1 ± 0.1 , 277 ± 2 and 75 ± 1 nmol $\text{kg}^{-1} \text{min}^{-1}$, respectively.

Calculations

From respiratory measurements, total fat and carbohydrate oxidation rates were calculated using the non-protein respiratory quotient [38]:

$$\text{Fat oxidation rate} = 1.695 V_{\text{O}_2} - 1.701 V_{\text{CO}_2} \quad (1)$$

$$\text{Carbohydrate oxidation rate} = 4.585 V_{\text{CO}_2} - 3.226 V_{\text{O}_2} \quad (2)$$

with V_{O_2} and V_{CO_2} in l/min and oxidation rates in g/min. Rate of appearance (R_a) and disappearance (R_d) of palmitate and glucose were calculated using the single-pool non-steady-state Steele equations adapted for stable isotope methodology [39]:

$$R_a = \frac{F - V[(C_2 + C_1)/2][(E_2 - E_1)/(t_2 - t_1)]}{(E_2 + E_1)/2} \quad (3)$$

$$R_d = R_a - V \cdot \left(\frac{C_2 - C_1}{t_2 - t_1} \right) \quad (4)$$

where F is the infusion rate ($\mu\text{mol kg}^{-1} \text{min}^{-1}$); V =distribution volume for palmitate or glucose (40 and 160 ml/kg, respectively); C_1 and C_2 are the palmitate or glucose concentrations (mmol/l) at time 1 (t_1) and 2 (t_2), respectively; and E_2 and E_1 are the plasma palmitate or glucose enrichments (TTR) at time 1 and 2, respectively. $^{13}\text{CO}_2$ production ($\text{Pr}^{13}\text{CO}_2$; mol/min) from the infused palmitate tracer was calculated as:

$$\text{Pr}^{13}\text{CO}_2 = (\text{TTR}_{\text{CO}_2} \cdot V_{\text{CO}_2}) / (k \cdot \text{Ar}) \quad (5)$$

where TTR_{CO_2} is the breath $^{13}\text{C}/^{12}\text{C}$ ratio at a given time point; V_{CO_2} is the carbon dioxide production (l/min); k is the volume of 1 mol of CO_2 (22.4 l/mol); and Ar is the fractional ^{13}C label recovery in breath CO_2 , observed after the infusion of labelled acetate [37] and calculated as:

$$\text{Ar} = (\text{TTR}_{\text{CO}_2} \cdot V_{\text{CO}_2}) / (k \cdot 2F) \quad (6)$$

where F is infusion rate of $[1,2-^{13}\text{C}]$ acetate (mol/min). Plasma palmitate oxidation rate (R_{ox}) (mol/min) can subsequently be calculated as:

$$R_{\text{ox}} \text{ palmitate} = R_d \text{ palmitate} (\text{Pr}^{13}\text{CO}_2 / F \cdot 16) \quad (7)$$

where R_d palmitate is the rate of disappearance of palmitate (mol/min); F is the palmitate infusion rate (mol/min); and 16 is the number of carbon atoms in palmitate. Plasma NEFA oxidation is calculated by dividing the plasma palmitate oxidation rate by the fractional contribution of palmitate to total NEFA concentration at each time point. Muscle (and lipoprotein)-derived TG oxidation is estimated by calculating the difference between plasma NEFA and total fat oxidation rate.

In a previous study [40] it has been shown that during exercise (50% W_{max}) plasma glucose R_d equals its R_{ox} (96–100%). Therefore, plasma glucose oxidation rate during exercise is calculated as:

$$R_{\text{ox}} \text{ plasma glucose} = R_d \text{ plasma glucose} \quad (8)$$

Muscle glycogen oxidation was calculated by subtracting plasma glucose oxidation from total carbohydrate oxidation. As plasma glucose R_d does not match R_{ox} during resting conditions, plasma glucose oxidation rates cannot be accurately calculated at rest and/or post-exercise recovery when using a $[6,6-^2\text{H}_2]$ glucose tracer [40].

Muscle sample analyses

Muscle samples were dissected, freed from any visible non-muscle material and frozen in nitrogen-cooled isopentane and embedded in Tissue-Tek (Sakura, Zoeterwoude, the Netherlands). Multiple serial sections (5 μm) were thaw-mounted together on uncoated, pre-cleaned glass slides for each subject. To determine muscle fibre type specific IMTG content, cross-sections were stained with oil red O together with immunolabelled cellular constituents using a previously described protocol [23, 25, 41]. For each muscle biopsy, a total of 84 ± 4 muscle fibres were analysed for lipid content. To permit quantification of intramyocellular glycogen, we used the modified PAS stain [42]. For each muscle biopsy 80 ± 4 muscle fibres were analysed for glycogen content. Large overviews containing 447 ± 31 fibres per subject were used to determine fibre type composition.

Statistics

All data are expressed as means \pm SEM. To compare tracer kinetics and/or plasma metabolite concentrations over time between trials, a two-way repeated measures analysis of variance was applied. A Scheffé's post hoc test was applied in case of a significant F -ratio to locate specific differences. For non-time-dependent variables, a two-tailed Student's t -test for paired observations was used. Significance was set at the 0.05 level of confidence.

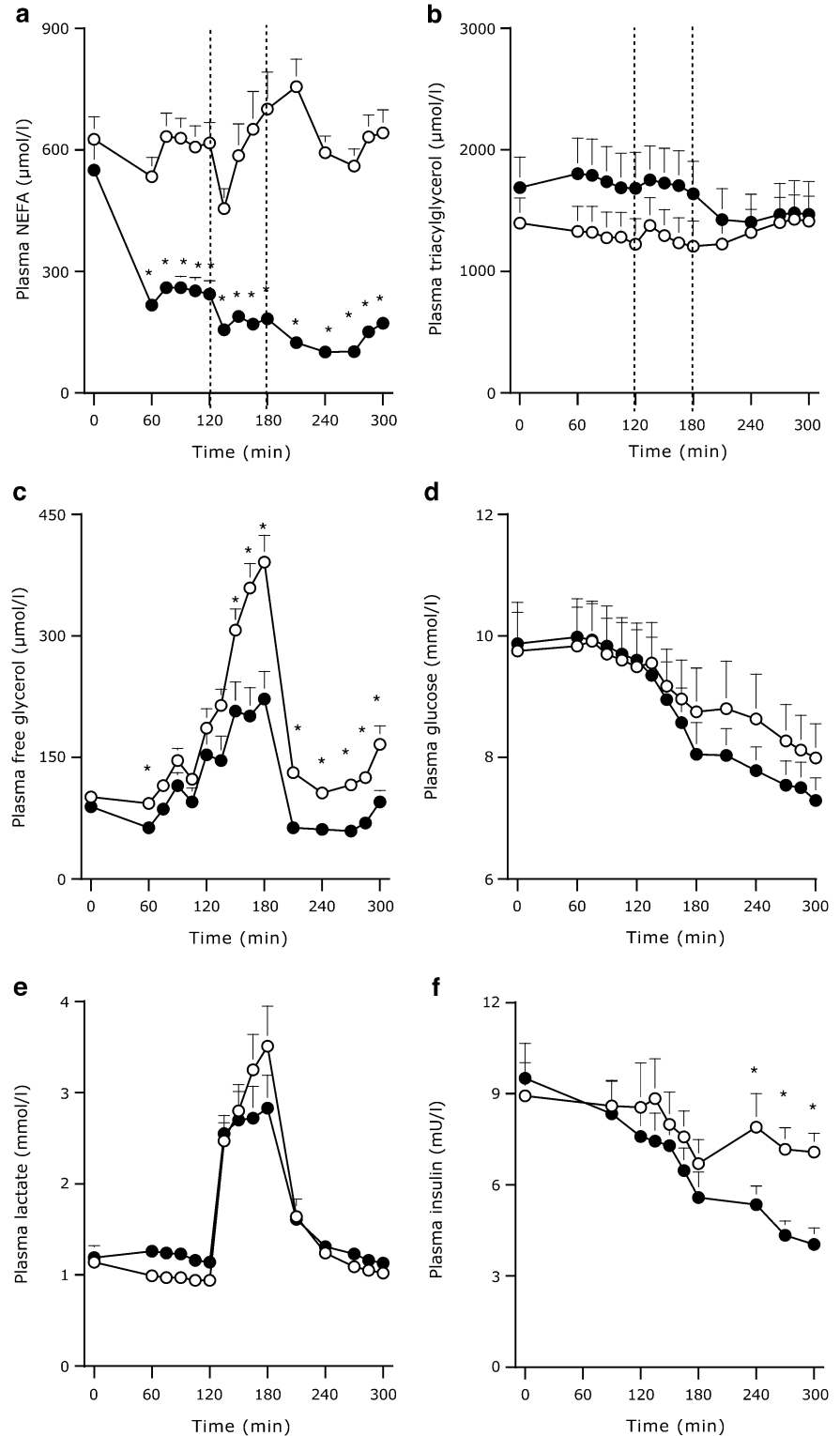
Results

Tracer kinetics

Plasma NEFA and glucose concentrations at rest, during exercise and recovery under normal, fasting conditions (CON) and reduced plasma NEFA availability (LFA) are

shown in Fig. 1. Plasma palmitate R_a , R_d and R_{ox} increased during exercise in the CON trial ($p < 0.01$), whereas in the LFA trial R_a and R_d remained stable (Fig. 2a). In both trials, palmitate R_a , R_d and R_{ox} were greater during exercise than resting values ($p < 0.05$; Table 2). At all stages, plasma palmitate R_a , R_d and R_{ox} were substantially lower in the LFA than in the CON trial ($p < 0.01$). Plasma glucose R_a

Fig. 1 Plasma NEFA, TG, glycerol, glucose, lactate and insulin concentrations at rest ($t=0-120$ min), during exercise ($t=120-180$ min) and post-exercise recovery ($t=180-300$ min) in both the CON (open circles) and LFA (closed circles) trial. Data represent means \pm SEM; * $p < 0.05$ for difference between trials



and R_d increased over time during exercise in both trials (Fig. 2b; $p<0.01$). In the CON trial, plasma glucose R_a was significantly higher during exercise than the resting values (Table 2). In both trials, plasma glucose R_d was significantly higher during exercise than the resting values

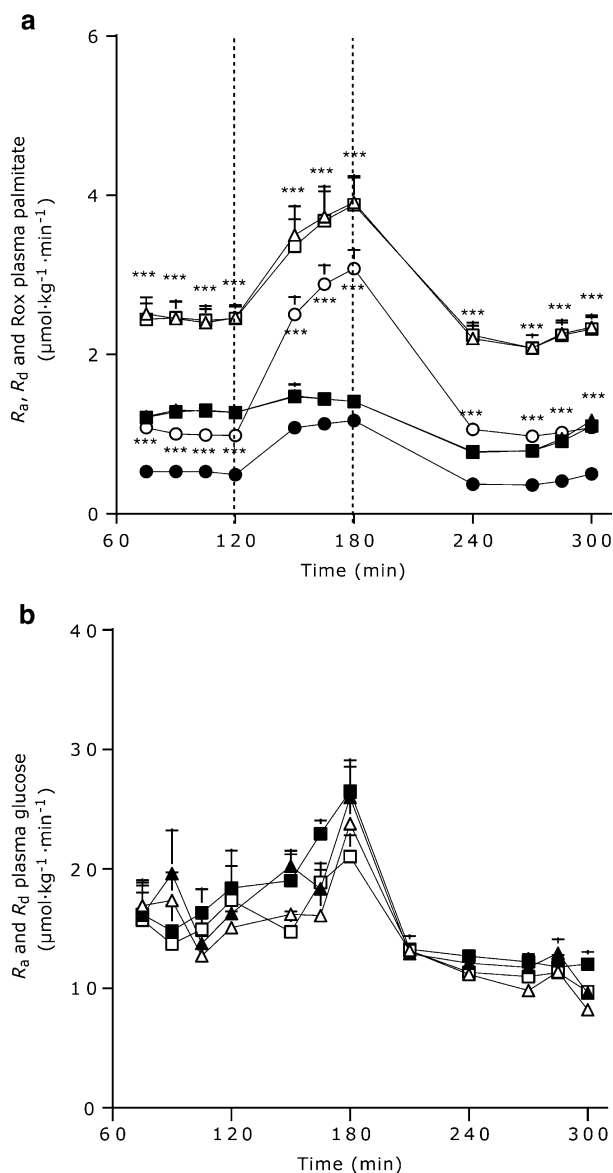


Fig. 2 Plasma palmitate rate of appearance (R_a), disappearance (R_d) and oxidation (R_{ox}) (a) and plasma glucose R_a and R_d (b) at rest ($t=60$ – 120 min), during exercise ($t=120$ – 180 min) and subsequent recovery ($t=180$ – 300 min) under normal, fasting conditions (white symbols; CON) and following administration of a peripheral lipolytic inhibitor (black symbols; LFA). Plasma palmitate R_a (open triangles), R_d (open squares) and R_{ox} (open circles) significantly increased during exercise in the CON trial ($p<0.001$), whereas in the LFA trial R_a and R_d remained stable over time. Palmitate R_a , R_d and R_{ox} were substantially lower at rest, during exercise and recovery in the LFA vs the CON trial ($p<0.001$). Plasma glucose R_a and R_d increased during exercise in both trials ($p<0.001$). Plasma palmitate R_a in the LFA trial (closed triangles) is nearly identical to palmitate R_d (closed squares) and symbols overlay. Data represent means \pm SEM; *** $p<0.001$ for difference in R_a , R_d and R_{ox} between trials

Table 2 Tracer kinetics at rest, during exercise and recovery

	Control trial (CON)	Low NEFA availability (LFA)
<i>Rest</i>		
R_a palmitate	2.46 \pm 0.18	1.27 \pm 0.12 ^a
R_d palmitate	2.45 \pm 0.18	1.27 \pm 0.11 ^a
R_{ox} palmitate	1.01 \pm 0.10	0.52 \pm 0.05 ^a
% R_{dox} palmitate	41.1 \pm 1.90	41.0 \pm 1.63
R_a glucose	15.5 \pm 0.8	15.4 \pm 1.4
R_d glucose	16.4 \pm 0.8	16.4 \pm 1.4
<i>Exercise</i>		
R_a palmitate	3.71 \pm 0.35 ^b	1.44 \pm 0.11 ^{ab}
R_d palmitate	3.64 \pm 0.34 ^b	1.44 \pm 0.11 ^{ab}
R_{ox} palmitate	2.82 \pm 0.22 ^b	1.13 \pm 0.10 ^{ab}
% R_{dox} palmitate	79.3 \pm 3.82 ^b	78.0 \pm 2.91 ^b
R_a glucose	18.7 \pm 1.0 ^b	18.2 \pm 1.1
R_d glucose	21.5 \pm 1.1 ^b	22.8 \pm 1.5 ^b
<i>Recovery</i>		
R_a palmitate	2.22 \pm 0.15	0.92 \pm 0.05 ^{ab}
R_d palmitate	2.22 \pm 0.15	0.89 \pm 0.04 ^{ab}
R_{ox} palmitate	1.03 \pm 0.08	0.41 \pm 0.03 ^{ab}
% R_{dox} palmitate	46.5 \pm 1.66 ^b	45.2 \pm 2.23 ^b
R_a glucose	10.1 \pm 0.5 ^b	10.8 \pm 0.4 ^b
R_d glucose	11.6 \pm 0.8 ^b	12.2 \pm 0.4 ^b

Tracer kinetics as calculated at rest ($t=90$ – 120 min), during exercise at 50% W_{max} ($t=120$ – 180 min) and during subsequent recovery ($t=240$ – 300 min)

Values are expressed as means \pm SEM ($n=10$)

R_a rate of appearance, R_d rate of disappearance, R_{ox} rate of oxidation ($\mu\text{mol kg}^{-1} \text{min}^{-1}$), % R_{dox} percentage of R_d palmitate oxidised (%)

^a $p<0.01$ for difference between trials

^b $p<0.05$ for difference from resting values

(Table 2). During recovery, glucose R_a and R_d were reduced compared with basal values in both trials ($p<0.05$). No differences in glucose kinetics were observed between trials.

Substrate utilisation

Energy expenditure and substrate use at rest are illustrated in Fig. 3a. Energy expenditure was similar between trials and averaged 5.75 \pm 0.22 and 5.76 \pm 0.27 kJ/min in the CON and LFA trial, respectively. However, substrate source utilisation rates differed significantly between trials ($p<0.05$). Total fat oxidation averaged 0.11 \pm 0.01 and 0.09 \pm 0.01 g/min, contributing 77 \pm 2 and 64 \pm 5% to total energy expenditure, in the CON and LFA trial, respectively ($p<0.05$). The difference in total fat oxidation was accounted for by a 38 \pm 6% reduction in plasma NEFA oxidation rate in the LFA compared to the CON trial (0.07 \pm 0.01 vs 0.11 \pm 0.01 g/min, respectively; $p<0.01$). In contrast, the use of TG sources was substantially augmented in the LFA trial (0.03 \pm 0.01 g/min), compared to the negligible IMTG use in the CON trial (0.00 \pm 0.01 g/min; $p<0.01$). Carbohydrate oxidation rates were significantly

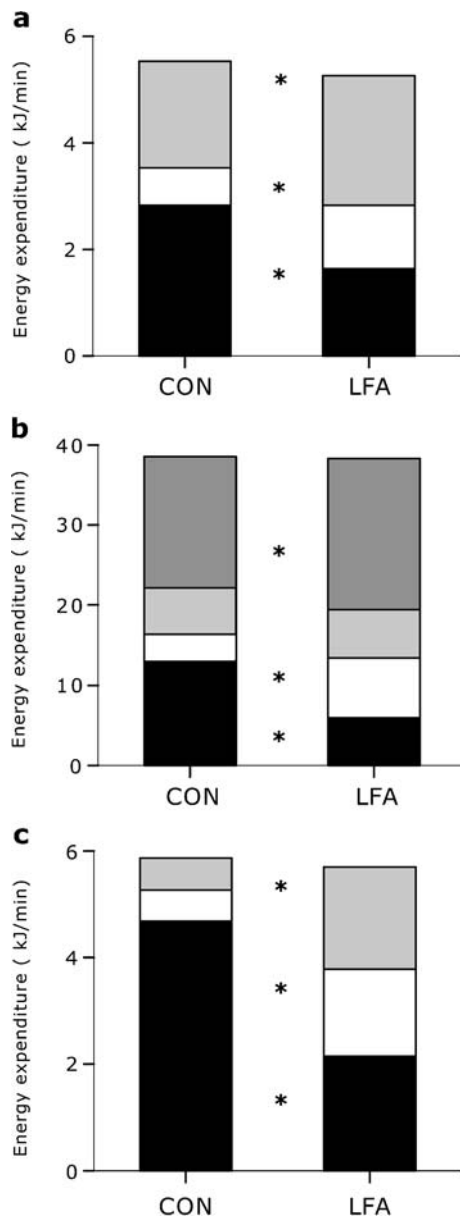


Fig. 3 Substrate source utilisation (in kJ/min) at rest ($t=90\text{--}120$ min) (a), during moderate intensity exercise ($t=120\text{--}180$ min) (b) and subsequent post-exercise recovery ($t=240\text{--}300$ min) (c) in the CON and LFA trial, respectively. Dark shading (a–c), plasma NEFA; white (a–c), muscle and lipoprotein-derived TG; pale grey (a, c), carbohydrate; pale grey (b), plasma glucose; mid-grey (b), muscle glycogen. * $p<0.05$ for differences in plasma NEFA, muscle (and lipoprotein)-derived TG, muscle glycogen and/or total carbohydrate oxidation rate between the CON and LFA trial

higher in the LFA vs the CON trial (0.13 ± 0.02 vs 0.08 ± 0.01 g/min, respectively; $p<0.01$).

In the subsequent exercise trial, the applied 50% W_{\max} workload averaged 100 ± 7 W ($63\pm2\%$ $V_{O_{2\max}}$). Substrate source utilisation rates are illustrated in Fig. 3b. Energy expenditure during exercise averaged 38.5 ± 2.2 and 38.3 ± 2.3 kJ/min in the CON and LFA trial, respectively. Total fat oxidation rate averaged 0.40 ± 0.04 and 0.33 ± 0.05 g/min, representing 42 ± 3 and $34\pm4\%$ of total energy expenditure

in the CON and LFA trial, respectively ($p<0.05$). In the CON trial, plasma NEFA oxidation provided $34\pm3\%$ of total energy expenditure, with TG sources contributing only $8\pm3\%$. In the LFA trial, plasma NEFA oxidation was substantially impaired, providing only $16\pm1\%$. In contrast, TG use was significantly augmented, providing as much as $18\pm5\%$ of total energy expenditure in the LFA trial. Total carbohydrate oxidation rates averaged 1.37 ± 0.09 and 1.54 ± 0.09 g/min in the CON and LFA trial, respectively ($p<0.01$). The greater endogenous carbohydrate use in the LFA trial was attributed to a greater reliance on muscle glycogen (0.98 ± 0.08 and 1.13 ± 0.10 g/min, respectively; $p<0.05$), as plasma glucose oxidation rates did not differ between trials (0.36 ± 0.03 and 0.37 ± 0.03 g/min, respectively).

Energy expenditure and substrate use during post-exercise recovery are illustrated in Fig. 3c. Energy expenditure was similar to pre-exercise resting values and averaged 5.86 ± 0.22 and 5.70 ± 0.26 kJ/min in the CON and LFA trial respectively. In the CON trial, fat oxidation rates were significantly greater during recovery than during pre-exercise rest (0.13 ± 0.01 vs 0.11 ± 0.01 g/min, respectively; $p<0.001$), which was predominantly attributed to a greater use of TG sources ($p=0.06$). The greater total fat oxidation rate during recovery was matched by reduced endogenous carbohydrate use (0.04 ± 0.01 vs 0.08 ± 0.01 g/min; $p<0.001$). In the LFA trial, fat and carbohydrate oxidation rates were similar during recovery compared to pre-exercise, resting values. However, during recovery plasma NEFA oxidation rates were substantially lower than pre-exercise resting values ($p<0.05$). The lower plasma NEFA oxidation rates were compensated by a greater IMTG use ($p=0.05$). Between trials, plasma NEFA oxidation rates were substantially lower in the LFA than in the CON trial (0.05 ± 0.004 vs 0.11 ± 0.008 g/min, respectively; $p<0.001$). These lower plasma NEFA oxidation rates were compensated by a greater muscle (and lipoprotein)-derived TG use (0.040 ± 0.009 vs 0.014 ± 0.006 g/min, respectively; $p<0.01$) as well as total endogenous carbohydrate use (0.12 ± 0.02 vs 0.04 ± 0.01 , respectively; $p<0.01$).

Plasma metabolite concentrations

Plasma metabolite concentrations are shown in Fig. 1. In the CON trial, plasma NEFA concentrations increased during exercise, reaching peak levels 15 min after cessation of exercise. Thereafter, NEFA concentrations declined to near pre-exercise resting levels. In contrast, in the LFA trial, plasma NEFA concentrations decreased after Acipimox administration and remained well below baseline values ($p<0.001$). Plasma TG gradually decreased during exercise in both trials ($p<0.001$). Plasma glycerol concentrations increased significantly during exercise and decreased during recovery in both trials ($p<0.001$). Glycerol concentrations were significantly lower in the LFA than in the CON trial ($p<0.001$). Plasma glucose concentra-

tions declined during exercise and recovery in both trials ($p<0.001$). Plasma glucose concentrations tended to decline more in the LFA trial ($p=0.08$). At $t=300$ min plasma glucose concentrations were significantly lower in the LFA trial ($p<0.05$). In both trials, plasma lactate concentrations increased well above pre-exercise resting levels during exercise, after which concentrations returned to near baseline values ($p<0.001$). Plasma insulin concentrations declined during exercise in both trials ($p<0.001$). During recovery, insulin levels continued to decline in the LFA trial only, resulting in significantly lower insulin concentrations in the LFA trial ($p<0.01$).

Muscle tissue analysis

Muscle fibre type analyses showed a $45\pm3\%$ type I and $55\pm3\%$ type II muscle fibre content. A significant decline of muscle fibre lipid content was observed in the LFA trial ($p<0.05$; Fig. 4). No changes in muscle lipid content were observed in the CON trial. The net decline in type I muscle fibre lipid content was significantly greater in the LFA than in the CON trial ($p<0.05$). Intramyocellular lipid content was ~three-fold greater in the type I than in type II fibres ($p<0.01$). Intramyocellular glycogen content declined substantially in both type I and II muscle fibres in both trials ($p<0.01$; Fig. 5). No differences in glycogen content were observed between trials or fibre types.

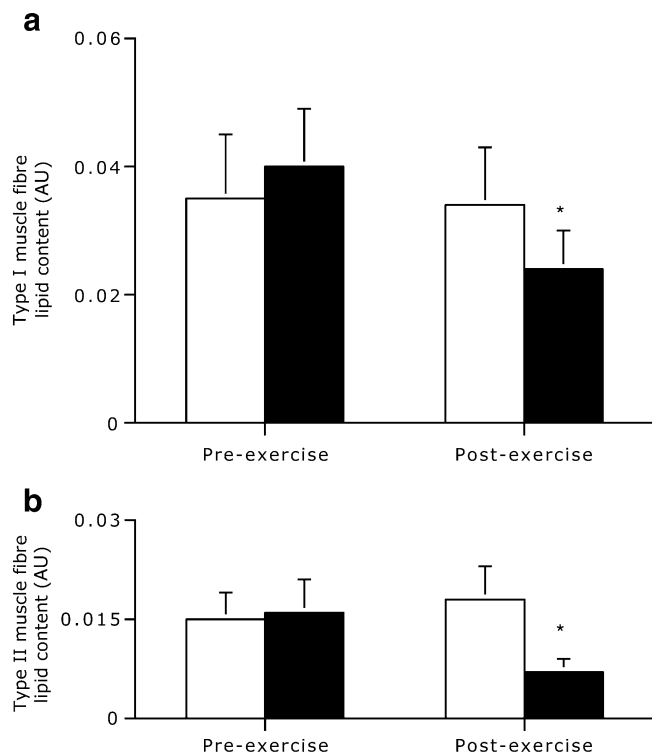


Fig. 4 Fibre-type-specific intramyocellular lipid content (% area lipid stained) before and immediately after exercise in the CON (white bars) and LFA (black bars) trial, as determined in type I (a) or type II (b) muscle fibres by using quantitative fluorescence microscopy on oil red O-stained muscle cross-sections. Data represent means \pm SEM; * $p<0.05$ for lower than pre-exercise values in each trial

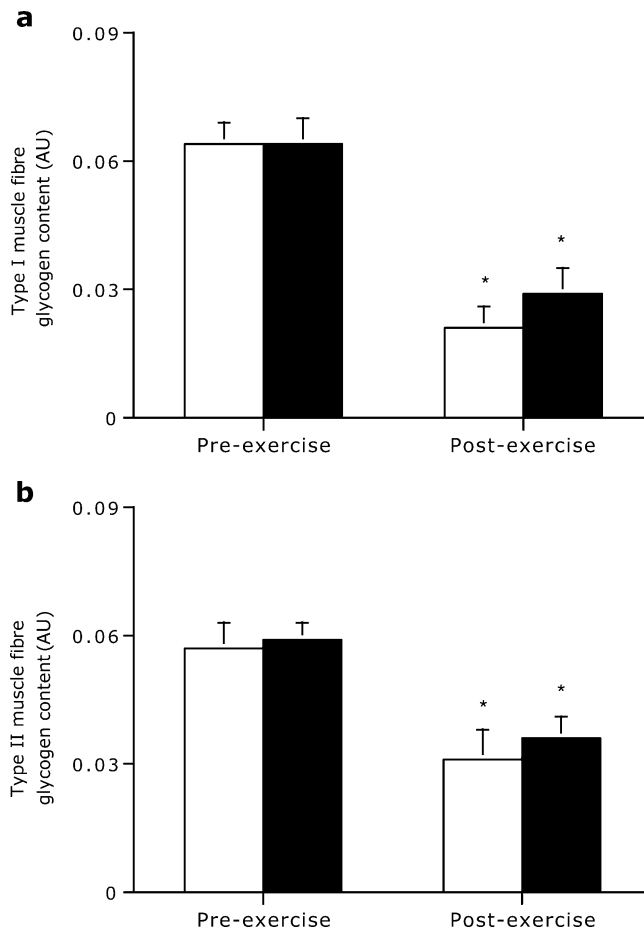


Fig. 5 Fibre-type-specific intramyocellular glycogen content (staining intensity) before and immediately after exercise in the CON (white bars) and LFA (black bars) trial, as determined in type I (a) or type II (b) muscle fibres by using the modified PAS stain method on muscle cross-sections. Data represent means \pm SEM; * $p<0.05$ for lower than pre-exercise values in each trial

Whole-body insulin sensitivity

The ratio between plasma glucose R_d and the plasma glucose and insulin concentrations was calculated as an index of whole-body insulin sensitivity. Changes in the insulin sensitivity index are illustrated in Fig. 6. During pre-exercise rest, insulin sensitivity tended to increase more in the LFA than in the CON trial ($p=0.088$; Fig. 6a). During exercise, the index increased progressively ($p<0.001$), with no differences between trials (Fig. 6b). During recovery, the index significantly increased in the LFA trial only, resulting in a significant difference between trials ($p<0.01$; Fig. 6c).

Discussion

In the present study, we show that Acipimox administration effectively inhibits adipose tissue lipolysis in vivo in type 2 diabetic patients, with plasma NEFA rate of appearance being decreased by a respective 46 ± 6 , 58 ± 4 and $57\pm3\%$ at rest, during exercise and post-exercise recovery

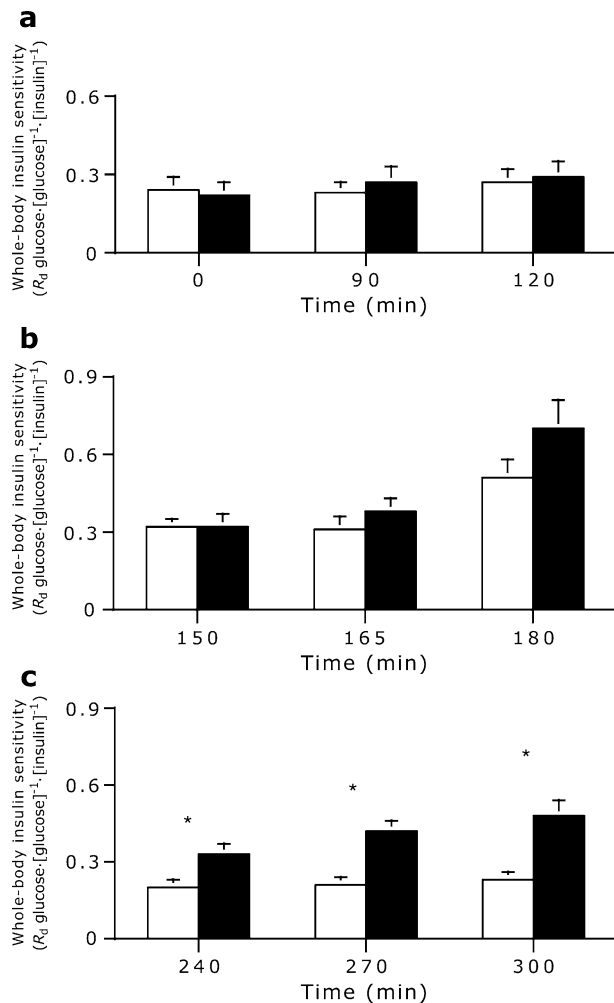


Fig. 6 Changes in whole-body insulin sensitivity (as determined by plasma glucose disappearance rate ($\mu\text{mol kg}^{-1} \text{min}^{-1}$) divided by the plasma glucose (mmol/l) and the plasma insulin concentration (mU/l) at rest (a), during exercise (b) and during post-exercise recovery (c) in the CON (white bars) and LFA (black bars) trial. Data represent means \pm SEM; * $p < 0.05$ for difference between trials

when compared to normal, fasting conditions ($p < 0.001$; Table 2, Figs. 1, 2). This substantially reduced systemic NEFA availability ($p < 0.001$), with plasma NEFA concentrations remaining well below baseline levels (Fig. 1). Plasma NEFA concentrations were lower at rest (~two-fold), during exercise (~three-fold) and recovery (~five-fold) in the LFA than in the CON trial ($p < 0.001$). From a quantitative point of view, IMTG does not represent an important substrate source under normal, fasting conditions in overweight type 2 diabetic patients. However, IMTG oxidation rates can be elevated substantially at rest, during exercise and subsequent recovery by reducing plasma NEFA availability (Figs. 3, 4, 5). This study shows that plasma NEFA availability regulates IMTG use. It also suggests that adipose tissue lipolytic inhibition could be an effective means of augmenting intramyocellular lipid use. The combination of exercise with adipose lipolytic inhibition seems to represent an effective strategy to aug-

ment post-exercise whole-body insulin sensitivity in type 2 diabetic patients in an overnight fasted state (Fig. 6).

There has been much controversy regarding the role of the IMTG pool as an available substrate source in humans [24, 43]. However, many of the inconsistent findings on the proposed metabolic importance of the IMTG pool can be attributed to methodological difficulties associated with the methods used to estimate IMTG use [43]. Recent studies applying isotope tracers, ^1H -magnetic-resonance spectroscopy, electron and/or immunofluorescence microscopy all support the contention that the IMTG pool functions as a readily available substrate source in healthy, lean men [24]. However, the role of the IMTG pool as an available substrate source in obese and/or type 2 diabetic patients remains to be elucidated [24]. In the present study, total fat oxidation at rest was entirely accounted for by plasma NEFA uptake and oxidation (CON trial). The contribution of fat sources other than plasma-derived NEFA was shown to be negligible (Fig. 3a). During exercise, the greater energy demands were matched by an increase in total fat and carbohydrate oxidation rates. Besides a three-fold increase in plasma NEFA oxidation rate (Table 2, Fig. 2a), other fat sources were mobilised to contribute to the greater energy demands (Fig. 3b). These other fat sources are generally assumed to reflect the use of muscle-derived TG. However, the applied methodology does not differentiate between muscle- and lipoprotein-derived TG use. Nonetheless, lipoprotein-derived TG oxidation seems to be of little quantitative importance in an overnight fasted state [24, 43]. In the present study, the contribution of muscle (and lipoprotein)-derived TG oxidation represented only $8 \pm 3\%$ of total energy expenditure during exercise (Fig. 3b). In accordance with this, we did not detect a significant net decline in intramyocellular lipid content in muscle samples collected before and after exercise (Fig. 4). Muscle glycogen oxidation contributed more substantially to total energy expenditure during exercise ($16 \pm 1\%$; Fig. 3b), resulting in a significant net decline in muscle glycogen content (Fig. 5). During recovery, muscle TG use tended to be greater compared with pre-exercise resting values ($p = 0.06$) and contributed $10 \pm 5\%$ of energy expenditure (Fig. 3c). Overall, these findings seem to be in line with previous reports suggesting that IMTG mobilisation and/or oxidation rates are reduced in obese and/or type 2 diabetic patients [24, 43]. However, direct comparisons of IMTG use at rest, during exercise and/or recovery between type 2 diabetic patients and matched, normoglycaemic controls are warranted to confirm those suggestions.

As elevated IMTG accretion in type 2 diabetic patients is the result of a structural imbalance between NEFA uptake, IMTG deposition and its subsequent mobilisation and/or oxidation [21–23], it has been suggested that an elevated IMTG turnover rate can prevent excessive accumulation of intramyocellular FA metabolites [9, 10, 13–16, 44, 45] and/or reduce the degree of intramyocellular lipid peroxidation [17]. In agreement, interventions known to

improve insulin sensitivity, like dietary lipid withdrawal [2], acute exercise [2, 25] and the use of thiazolidinediones [46] have all been associated with a reduction in skeletal muscle lipid content. Recently, we as well as others [31], have shown that pharmacological inhibition of adipose tissue lipolysis can prevent the downregulation of IMTG use with time during prolonged exercise, which was shown to be associated with the concomitant increase in plasma NEFA concentrations. Subsequently, we speculated whether the same intervention could also effectively lower plasma NEFA availability in type 2 diabetic patients and increase the capacity to mobilise and oxidise the IMTG stores at rest and/or during exercise. Therefore, we administered a nicotinic acid analogue to specifically inhibit adipose tissue lipolysis [47, 48]. In response, plasma NEFA R_a , R_d and R_{ox} were substantially reduced (Table 2, Figs. 1, 2, 3), while muscle TG and endogenous carbohydrate oxidation rates were significantly elevated (Fig. 3). Accordingly, a significant decline in intramyocellular lipid content was observed following exercise in the LFA trial only (Fig. 4). The greater carbohydrate use during exercise in the LFA trial was attributed to an elevated muscle glycogen utilisation rate. However, histological analyses did not show a greater decline in intramyocellular glycogen content in the LFA vs the CON trial (Fig. 5). In contrast, Galbo et al. [33] reported a greater reduction in muscle glycogen content following exercise with nicotinic acid administration. In the present study, the apparent discrepancy between the stable isotope data and muscle sample analysis is probably explained by the heterogeneity of muscle biopsy samples, the sensitivity of the histological staining technique and/or the relative short duration of the exercise trial.

The depletion of the muscle lipid and/or glycogen stores has been associated with exercise-induced increase in skeletal muscle insulin sensitivity [24, 49–51]. Consequently, it could be speculated that exercise interventions that augment IMTG and/or glycogen use are more effective in improving insulin sensitivity. Furthermore, peroxisome proliferator-activated receptor gamma agonists, like thiazolidinediones, improve insulin sensitivity partly through enhanced lipid retention in adipose tissue, thereby stimulating the redistribution of TG from skeletal muscle and liver back to adipose tissue [46]. Nicotinic acid analogues are likely to act along the same paradigm. Though we did not aim to assess the effects of exercise and/or acute adipose tissue lipolytic inhibition on insulin sensitivity, some interesting findings were reported. Hepatic glucose output (R_a) and glucose disposal (R_d) rates were significantly lower during recovery when compared to pre-exercise resting values (Table 2). Though we observed no differences in glucose disposal between trials, insulin concentrations significantly declined in the LFA trial only (Fig. 1). After calculating the ratio between plasma glucose R_d and plasma glucose and insulin concentrations as an index of whole-body insulin sensitivity, we observed a significant increase in insulin sensitivity during recovery in the LFA trial (Fig. 6c). The latter seems to support our hypothesis that exercise, in combination

with acute lipolytic inhibition, can augment the exercise-induced increase in skeletal muscle insulin sensitivity. More research is warranted to investigate this relationship and its potential clinical relevance.

In conclusion, administration of a nicotinic acid analogue effectively suppresses adipose tissue lipolysis, both at rest, during exercise and subsequent recovery in an overnight fasted state. The reduction in plasma NEFA availability stimulates intramuscular lipid and glycogen use at rest, during exercise and/or subsequent recovery. Inhibition of adipose tissue lipolysis during exercise provides an effective mechanistic approach to significantly reduce intramyocellular lipid deposits. The latter could represent an effective interventional strategy to improve skeletal muscle insulin sensitivity in type 2 diabetic patients.

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References

- Oakes ND, Cooney GJ, Camilleri S, Chisholm DJ, Kraegen EW (1997) Mechanisms of liver and muscle insulin resistance induced by chronic high-fat feeding. *Diabetes* 46:1768–1774
- Oakes ND, Bell KS, Furler SM et al (1997) Diet-induced muscle insulin resistance in rats is ameliorated by acute dietary lipid withdrawal or a single bout of exercise: parallel relationship between insulin stimulation of glucose uptake and suppression of long-chain fatty acyl-CoA. *Diabetes* 46:2022–2028
- Storlien LH, Jenkins AB, Chisholm DJ, Pascoe WS, Khouri S, Kraegen EW (1991) Influence of dietary fat composition on development of insulin resistance in rats. Relationship to muscle triglyceride and omega-3 fatty acids in muscle phospholipid. *Diabetes* 40:280–289
- Pan DA, Lillioja S, Kriketos AD et al (1997) Skeletal muscle triglyceride levels are inversely related to insulin action. *Diabetes* 46:983–988
- Perseghin G, Scifo P, De Cobelli F et al (1999) Intramyocellular triglyceride content is a determinant of in vivo insulin resistance in humans: a ^1H - ^{13}C nuclear magnetic resonance spectroscopy assessment in offspring of type 2 diabetic parents. *Diabetes* 48:1600–1606
- Jacob S, Machann J, Rett K et al (1999) Association of increased intramyocellular lipid content with insulin resistance in lean nondiabetic offspring of type 2 diabetic subjects. *Diabetes* 48:1113–1119
- Shulman GI (1999) Cellular mechanisms of insulin resistance in humans. *Am J Cardiol* 84:3J–10J
- Randle PJ, Garland PB, Hales CN, Newsholme EA (1963) The glucose–fatty acid cycle: its role in insulin sensitivity and the metabolic disturbances of diabetes mellitus. *Lancet* 1:785–789
- Yu C, Chen Y, Cline GW et al (2002) Mechanism by which fatty acids inhibit insulin activation of insulin receptor substrate-1 (IRS-1)-associated phosphatidylinositol 3-kinase activity in muscle. *J Biol Chem* 277:50230–50236
- Roden M, Price TB, Perseghin G et al (1996) Mechanism of free fatty acid-induced insulin resistance in humans. *J Clin Invest* 97:2859–2865

11. Griffin ME, Marcucci MJ, Cline GW et al (1999) Free fatty acid-induced insulin resistance is associated with activation of protein kinase C θ and alterations in the insulin signaling cascade. *Diabetes* 48:1270–1274
12. Boden G, Lebed B, Schatz M, Homko C, Lemieux S (2001) Effects of acute changes of plasma free fatty acids on intramyocellular fat content and insulin resistance in healthy subjects. *Diabetes* 50:1612–1617
13. Shulman GI (2000) Cellular mechanisms of insulin resistance. *J Clin Invest* 106:171–176
14. Adams JM, Pratipanawatr T, Berria R et al (2004) Ceramide content is increased in skeletal muscle from obese insulin-resistant humans. *Diabetes* 53:25–31
15. Ellis BA, Poynten A, Lowy AJ et al (2000) Long-chain acyl-CoA esters as indicators of lipid metabolism and insulin sensitivity in rat and human muscle. *Am J Physiol* 279:E554–E560
16. Itani SI, Ruderman NB, Schmieder F, Boden G (2002) Lipid-induced insulin resistance in human muscle is associated with changes in diacylglycerol, protein kinase C, and I κ B- α . *Diabetes* 51:2005–2011
17. Russell AP, Gastaldi G, Bobbioni-Harsch E et al (2003) Lipid peroxidation in skeletal muscle of obese as compared to endurance-trained humans: a case of good vs. bad lipids? *FEBS Lett* 551:104–106
18. Rasmussen BB, Holmback UC, Volpi E, Morio-Liondore B, Paddon-Jones D, Wolfe RR (2002) Malonyl coenzyme A and the regulation of functional carnitine palmitoyl-transferase-1 activity and fat oxidation in human skeletal muscle. *J Clin Invest* 110:1687–1693
19. Jensen MD (2002) Fatty acid oxidation in human skeletal muscle. *J Clin Invest* 110:1607–1609
20. Kelley DE, He J, Menshikova EV, Ritov VB (2002) Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. *Diabetes* 51:2944–2950
21. Goodpaster B, Kelley D, Wing R, Meier A, Thaete F (1999) Effects of weight loss on regional fat distribution and insulin sensitivity in obesity. *Diabetes* 48:839–847
22. Thamer C, Machann J, Bachmann O et al (2003) Intramyocellular lipids: anthropometric determinants and relationships with maximal aerobic capacity and insulin sensitivity. *J Clin Endocrinol Metab* 88:1785–1791
23. van Loon LJC, Koopman R, Manders R, van der Weegen W, van Kranenburg GP, Keizer HA (2004) Intramyocellular lipid content in type 2 diabetes patients compared to overweight sedentary men and highly trained endurance athletes. *Am J Physiol* 287:E558–E565
24. van Loon LJC (2004) Use of intramuscular triacylglycerol as a substrate source during exercise in humans. *J Appl Physiol* 97:1170–1187
25. van Loon LJC, Koopman R, Stegen JH, Wagenmakers AJ, Keizer HA, Saris WH (2003) Intramyocellular lipids form an important substrate source during moderate intensity exercise in endurance-trained males in a fasted state. *J Physiol* 553:611–625
26. Watt MJ, Heigenhauser GJ, Dyck DJ, Spriet LL (2002) Intramuscular triacylglycerol, glycogen and acetyl group metabolism during 4 h of moderate exercise in man. *J Physiol* 541:969–978
27. Romijn JA, Coyle EF, Sidossis LS et al (1993) Regulation of endogenous fat and carbohydrate metabolism in relation to exercise intensity and duration. *Am J Physiol* 265:E380–E391
28. Romijn JA, Coyle EF, Sidossis LS, Zhang XJ, Wolfe RR (1995) Relationship between fatty acid delivery and fatty acid oxidation during strenuous exercise. *J Appl Physiol* 79:1939–1945
29. Reaven GM, Hollenbeck C, Jeng CY, Wu MS, Chen YD (1988) Measurement of plasma glucose, free fatty acid, lactate, and insulin for 24 h in patients with NIDDM. *Diabetes* 37:1020–1024
30. van Loon LJC, Thomason-Hughes M, Constantin-Teodosiu D et al (2005) Inhibition of lipolysis increases intramuscular lipid and glycogen use in vivo in humans. *Am J Physiol Endocrinol Metab* DOI 10.1152/ajpendo.00092.2005
31. Watt MJ, Holmes AG, Steinberg GR, Mesa JL, Kemp BE, Febbraio MA (2004) Reduced plasma FFA availability increases net triacylglycerol degradation, but not GPAT or HSL activity, in human skeletal muscle. *Am J Physiol* 287:E120–E127
32. Issekutz B Jr, Paul P (1968) Intramuscular energy sources in exercising normal and pancreatectomized dogs. *Am J Physiol* 215:197–204
33. Galbo H, Holst JJ, Christensen NJ, Hilsted J (1976) Glucagon and plasma catecholamines during beta-receptor blockade in exercising man. *J Appl Physiol* 40:855–863
34. Alberti KG, Zimmet PZ (1998) Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. *Diabet Med* 15:539–553
35. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC (1985) Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 28:412–419
36. Siri WE (1956) The gross composition of the body. *Adv Biol Med Physiol* 4:238–280
37. van Loon LJC, Koopman R, Schrauwen P, Stegen J, Wagenmakers AJ (2003) The use of the [1,2- 13 C]acetate recovery factor in metabolic research. *Eur J Appl Physiol* 89:377–383
38. Peronnet F, Massicotte D (1991) Table of nonprotein respiratory quotient: an update. *Can J Sport Sci* 16:23–29
39. Wolfe RR, Jahoor F (1990) Recovery of labeled CO $_2$ during the infusion of C-1- vs C-2-labeled acetate: implications for tracer studies of substrate oxidation. *Am J Clin Nutr* 51:248–252
40. Jeukendrup AE, Raben A, Gijzen A et al (1999) Glucose kinetics during prolonged exercise in highly trained human subjects: effect of glucose ingestion. *J Physiol* 515:579–589
41. van Loon LJC, Schrauwen-Hinderling VB, Koopman R et al (2003) Influence of prolonged endurance cycling and recovery diet on intramuscular triglyceride content in humans. *Am J Physiol* 553:611–625
42. Schaart G, Hesselink RP, Keizer HA, Van Kranenburg G, Drost MR, Hesselink MK (2004) A modified PAS stain combined with immunofluorescence for quantitative analyses of glycogen in muscle sections. *Histochem Cell Biol* 122:161–169
43. Watt MJ, Heigenhauser GJ, Spriet LL (2002) Intramuscular triacylglycerol utilization in human skeletal muscle during exercise: is there a controversy? *J Appl Physiol* 93:1185–1195
44. Boden G, Shulman GI (2002) Free fatty acids in obesity and type 2 diabetes: defining their role in the development of insulin resistance and beta-cell dysfunction. *Eur J Clin Invest* 32 (S3):14–23
45. Dresner A, Laurent D, Marcucci M et al (1999) Effects of free fatty acids on glucose transport and IRS-1-associated phosphatidylinositol 3-kinase activity. *J Clin Invest* 103:253–259
46. Oakes ND, Camilleri S, Furler SM, Chisholm DJ, Kraegen EW (1997) The insulin sensitizer, BRL 49653, reduces systemic fatty acid supply and utilization and tissue lipid availability in the rat. *Metabolism* 46:935–942
47. Tunaru S, Kero J, Schaub A et al (2003) PUMA-G and HM74 are receptors for nicotinic acid and mediate its anti-lipolytic effect. *Nat Med* 9:352–355
48. Christie AW, McCormick DK, Emmison N, Kraemer FB, Alberti KG, Yeaman SJ (1996) Mechanism of anti-lipolytic action of acipimox in isolated rat adipocytes. *Diabetologia* 39:45–53
49. Perseghin G, Price TB, Petersen KF et al (1996) Increased glucose transport, phosphorylation and muscle glycogen synthesis after exercise training in insulin-resistant subjects. *N Engl J Med* 335:1357–1362
50. Wojtaszewski JF, Hansen BF, Kiens B, Richter EA (1997) Insulin signaling in human skeletal muscle: time course and effect of exercise. *Diabetes* 46:1775–1781
51. Wojtaszewski JF, Hansen BF, Gade et al (2000) Insulin signaling and insulin sensitivity after exercise in human skeletal muscle. *Diabetes* 49:325–331